OXFORD LAB FINE CHEM LLP
ISO 9001-2008 Certified Company

Regd Office: Unit no 12, 1st Floor,
Neminath Industrial Estate No.6,
Navghar, Vasai (East), Palghar - 410210.
Maharashtra, INDIA.

Tel: +91 250 2390032 / 2390989 / 2390990
Email: sales@oxfordlabchem.com /
info@oxfordlabchem.com
Web: www.oxfordlabchem.com

Oxford
Range of
Laboratory Chemicals

TECHNICAL DATA SHEET

Haemophilus Test Agar Base

Principle

Haemophilus Test Agar Base is intended for use in the antimicrobial disc diffusion susceptibility testing of *Haemophilus* species with supplements of either X-factor (hemin) or V-factor (nicotinamide adenine dinucleotide, NAD) and yeast extract. Haemophilus Test Agar Base is used for *Haemophilus* species as described in the Clinical and Laboratory Standards Institute (CLSI) for both dilution and disc diffusion assays. Media is composed of meat infusion form (equivalent of beef infusion form), casamino acids, starch, yeast extract, and agar. Meat infusion from and casamino acids provides nitrogen vitamins, amino acids, minerals, carbon, and other nutrients to support the growth of microorganisms. Yeast extract serves as a source of a vitamin B complex. While starch absorbs any toxic metabolites produced during microbial growth and while autoclaving undergoes hydrolysis and liberates small amounts of dextrose, which acts as a source of energy. Agar is the solidifying agent.

Use: For the susceptibility testing of Haemophilus influenza.

Contents*

Ingredients	Gram/Liter
Meat Infusion Form 300g#	2.000
Casamino Acids	17.500
Starch	1.500
Yeast Extract	5.000
Agar	17.000
pH at 25˚C	7.4 ± 0.2

*** Formula adjusted for optimum performance and parameters**
 # Equivalent to Beef infusion form

Directions: Dissolve 43.0 grams in 1000 ml distilled water. Boil to dissolve the medium completely and sterilize by autoclaving at 15 lbs. pressure (121 °C) for 15 min, cool it to 42-45 °C and aseptically add 15 mg NAD and 15 mg hematin in to the medium and mix well. Distribute aseptically in petri plates. Ensure complete solidification and inoculate test sample aseptically.

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Specimens' types analyzed
Isolated microorganism from non-clinical and clinical samples etc.

Precautions to be taken
These microbial media are intended for the in-vitro use only. All the handling, experiments, storage, and discarding should be performed with the help of skilled and knowledgeable technicians and as per the established guidelines. The material should be disposed only after proper sterilization by autoclaving. Please go through the MSDS of the media to avoid any accidents or in emergency.

Performance and Evaluation
The expected performance of the medium is liable to use as per the direction on the label when stored at optimum conditions and within expiry date.

Quality Control

Appearance	**Beige colored free flowing, homogeneous powder**
Reaction of 4.3% solution	**7.4 ± 0.2 at 25 ˚C**
pH	**7.20- 7.60**
Gelling	**Firm comparable with 1.7% agar gel**
Color and clarity of ready medium	**Light amber opalescent gel**
Growth Promotion properties	**Best at ≤ 100 CFU at 30-37 ºC for 18-72 h**
Indicative properties	**Optimum at ≤ 100 CFU at 30-37 ºC for 18-48 h**
Negative control	**Performed using sterile distilled water**

Different Microbial Response
Cultural characteristics observed after incubation at 35-37°C for 18-48 hours.

Organism	ATCC	Inoculum	Growth
Escherichia coli	**8739**	**50-100**	**Luxurious**
Salmonella typhi	**14028**	**50-100**	**Fair- Good**
Bacillus spizizenii	**6633**	**50-100**	**Luxurious**
Staphylococcus aureus	**25923**	**50-100**	**Luxurious**

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Storage and Shelf Life: **The product is highly hygroscopic; keep the container tightly closed at all times and store it properly as per the conditions mentioned on the label. The declared expiry is valid only when stored as per the conditions mentioned on the label.**
Note: Sterilize media immediately after reconstitution.

Disposal: **To avoid the contamination or propagation of any hazardous microbes the used, unusable or modified preparation of this product must be disposed after autoclaving after completion of task.**

Reference

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2. **Difco Manual (1998). 11th Edition. Difco Laboratories., Division of Becton Dickinson and Company, Sparks, Maryland, USA.**
3. **Isenberg, H.D. Clinical Microbiology Procedures Handbook 2nd Edition.**
4. **Jorgensen, Redding, Maher and Howell. 1987. J. Clin. Microbiol. 25:2105-2113.**
5. **Jorgensen, Turnidge and Washington. 1999. In Murray, Baron, Pfaller, Tenover and Yolken (ed.), Manual of clinical microbiology, 7th ed. American Society for Microbiology, Washington, D.C.**
6. **Mueller, J. H., and J. Hinton (1941). *A protein-free medium for primary isolation of gonococcus and meningococcus.* Proc. Soc. Exp. Biol. Med. 48:330-333.**

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