

## TECHNICAL DATA SHEET

### Rose Bengal Chloramphenicol Agar

#### Principle

Rose Bengal chloramphenicol agar is composed of mycological peptone, dextrose, potassium dihydrogen phosphate, magnesium sulphate, rose Bengal, chloramphenicol and agar. Mycological peptone provides carbon, nitrogen substances, long chain amino acids, vitamins and other essential growth nutrients. Dextrose is an energy source. Monopotassium phosphate acts as buffering agent. Magnesium sulfate provides ions for metabolic reactions. Rose Bengal is a selective agent, inhibits bacterial growth and prevent over growth of fastidious fungi and aids in isolation of slow growing fungi. It also reduces the spreading of molds, controls the size and height of molds colonies such as Rhizopus species. The rose Bengal is absorbed by the yeast and molds and help in identification and enumeration. Agar is solidifying agent. Chloramphenicol has inhibitory action on gram-negative bacteria.

**Use:** For selective isolation & enumeration of yeasts & molds from environmental materials & foods.

#### Contents\*

Ingredients	Gram/Liter
Mycological peptone	5.00
Dextrose	10.00
Potassium dihydrogen phosphate	1.00
Magnesium sulphate	0.50
Rose bengal	0.05
Chloramphenicol	0.10
Agar	15.00
pH at 25°C	7.2 ±0.2

\* Formula adjusted for optimum performance and parameters

**Directions:** Dissolve 32.00 grams in 1000 ml distilled water. Boil to dissolve the medium completely and sterilize by autoclaving at 15 lbs pressure (121°C) for 15 min, cool it to 42-45 °C and distribute aseptically in petri plates. Ensure complete solidification and inoculate test sample aseptically.

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## Specimens types analyzed

Food and dairy samples, environmental samples etc.

## Precautions to be taken

These plant tissue culture media are intended for the in-vitro use only. All the handling, experiments, storage, and discarding should be performed with the help of skilled and knowledgeable technicians and as per the established guidelines. The material should be disposed only after proper sterilization by autoclaving. Please go through the MSDS of the media to avoid any accidents or in emergency.

## Performance and Evaluation

The expected performance of the medium is liable to use as per the direction on the label when stored at optimum conditions and within expiry date.

## Quality Control

<b>Appearance</b>	<b>Pinkish beige colored free flowing, homogeneous powder</b>
<b>Reaction of 3.2% solution</b>	<b>7.2 ±0.2 at 25 °C</b>
<b>pH</b>	<b>7.00- 7.40</b>
<b>Gelling</b>	<b>Firm comparable with 1.5% agar gel</b>
<b>Color and clarity of ready medium</b>	<b>Pink colored opalescent gel</b>
<b>Growth Promotion properties</b>	<b>Best at ≤ 100 CFU at 25-30°C for 2-7 days</b>
<b>Indicative properties</b>	<b>Optimum at ≤ 100 CFU at 25-30°C for 3-5 days</b>
<b>Negative control</b>	<b>Performed using sterile distilled water</b>

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**Different Microbial Response: Cultural characteristics observed after incubation at 25-30 for 3-7 days. Inoculum 50-100 CFU.**

Organism	ATCC	Inoculum (CFU)	Growth	Recovery
<i>Candida albicans</i>	10231	50-100	Luxuriant	≥ 60%
<i>Saccharomyces cerevisiae</i>	9763	50-100	Luxuriant	≥ 60%
<i>Aspergillus brasiliensis</i>	16404	50-100	Luxuriant	≥ 60%
<i>Escherichia coli</i>	8739	50-100	Inhibited	--
<i>Bacillus subtilis</i>	6633	50-100	Inhibited	--

**Storage and Shelf Life:** The product is highly hygroscopic; keep the container tightly closed at all times and store it properly as per the conditions mentioned on the label. The declared expiry is valid only when stored as per the conditions mentioned on the label. Note: Sterilize media immediately after reconstitution.

**Disposal:** To avoid the contamination or propagation of any hazardous microbes the used, unusable or modified preparation of this product must be disposed after autoclaving after completion of task.

## Reference

1. Atlas, R. M. (2005). Handbook of media for environmental microbiology. CRC press.
2. Difco Manual (1998). 11<sup>th</sup> Edition. Difco Laboratories., Division of Becton Dickinson and Company, Sparks, Maryland, USA.
3. Murray P. R., Baron J. H., Pfaller M. A., Jorgensen J. H. and Tenover F. C., (Ed.), 2003, Manual of Clinical Microbiology, 8th Ed., American Society for Microbiology, Washington, D. C.

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